

An integrated Bayesian model for genotyping and copy number data



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Summary
• We derive a new method for the joint estimation of CN events and IBD/UPD regions
• It takes into account all errors in the microarray genotyping measurements, due to CN aberrations
• The goodness of our model is supported by the results on real data

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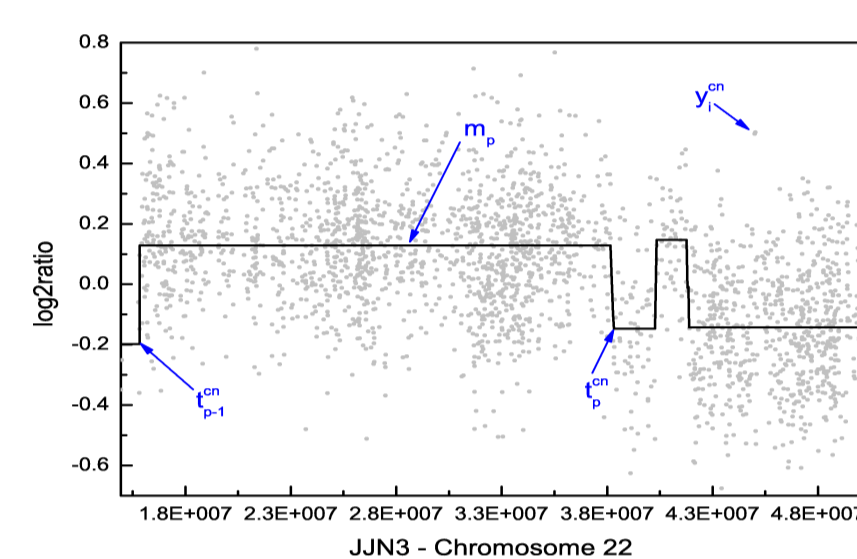
Genotyping and copy number data

- Single nucleotide polymorphism (SNPs) = single base-pair location in the genome where the nucleotide can assume two possible values among the four bases (T, A, C, G)
- We have two copies of each chromosome ⇒ at each SNP corresponds a pair of nucleotides:

AB } Heterozygosity or Het
 AA } Homozygosity or Hom
 BB }

where A and B are the two possible values of the SNP

- DNA copy number (CN) = for a given genomic region, is the number of copies of DNA of that region (normal CN = 2) ⇒ we can divide the genome in regions of constant CN, i.e. is a piecewise constant function of k^{cn} intervals with boundaries $t_0^{cn} = 0 = t_0^{cn}, t_1^{cn}, \dots, t_{k_0}^{cn} = n$ and levels of the segments $m \in \mathbb{R}^{k^{cn}}$ (usually a \log_2 ratio scale is used)



- Type of aberrations regarding genotyping and copy number data:

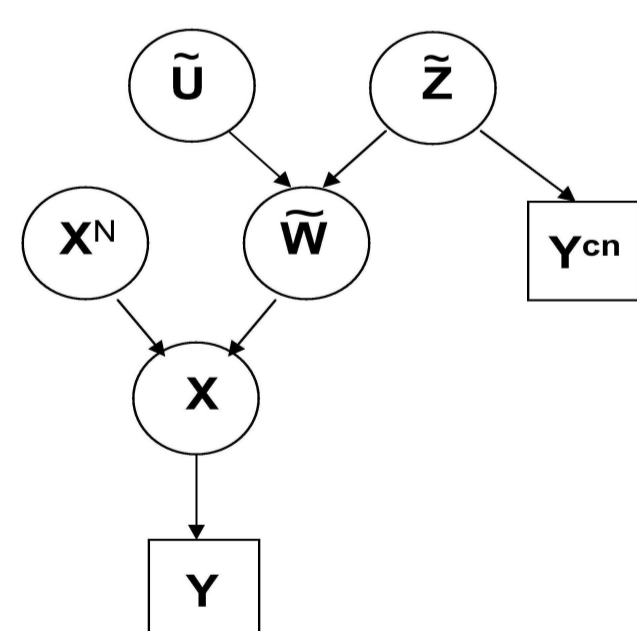
- amplification (CN > 4) ⇒ $\{Z = 2\}$
- gain (CN = 3, 4) ⇒ $\{Z = 1\}$
- loss (CN = 1) ⇒ $\{Z = -1\}$
- homozygous deletion (CN = 0) ⇒ $\{Z = -2\}$
- Loss of heterozygosity (LOH) with normal copy number, i.e. unusual long stretches of homozygous SNPs due to uniparental disomy or autozygosity (called IBD/UPD regions)

where Z is the r.v. which represents the CN aberration occurred ($\{Z = 0\}$ is the normal CN)

SNP microarray

- SNP microarrays are able to measure simultaneously genotyping and copy number data
- Microarray technology is not able to distinguish between the loss of one allele (e.g. A) or an Homozygosity (e.g. AA)
⇒ Integration of the two types of data to better identifies the aberrations (e.g. it possible to distinguish between IBD/UPD and loss or between gain and high amplification)
⇒ Bayesian regression to estimate the piecewise constant profile of the aberrations $\tilde{W} = (\tilde{W}_1, \dots, \tilde{W}_n)$ at n SNP loci. The profile consists of k_0 intervals, with boundaries $0 = t_0^0 < t_1^0 < \dots < t_{k_0-1}^0 < t_{k_0}^0 = n$, so that $\tilde{W}_{t_{p-1}^0+1} = \dots = \tilde{W}_{t_p^0} =: W_p$, for all $p = 1, \dots, k_0$.

The model



\underline{Y} = vector of the SNP genotypes detected by the microarray ($Y_i \in \mathbb{Y}$), where $\mathbb{Y} = \{Het, NHet, NoCall\}$ and $NHet$ = not Het

\underline{X} = vector of the true SNP genotypes in cancer cells ($X_i \in \mathbb{X}$), where $\mathbb{X} = \{Het, Hom\}$

\underline{X}^N = vector of the true SNP genotypes in normal cells ($X_i^N \in \mathbb{X}$)

\underline{Z} = vector of the CN aberrations

\underline{U} = vector of the occurrence of IBD/UPD

\underline{Y}^{cn} = vector of the raw CN data

⇒ for each interval p , $\{W_p = w\} = \{Z_p = z, U_p = u\}$

$P(\tilde{y}_i | \tilde{w}_i, x_i^N)$ estimated on two public datasets (Zhao et al. (2004), Forconi et al. (2008))

The priors & the posterior

The priors are defined as following:

- $P(X_i^N = Het)$ set on the basis of the microarray annotation file
- for $P(\tilde{U}_i = 1)$, we tried two values 0.001 and 0.0001, on the basis of the estimations obtained using the data in Bacolod et al. (2008) and The International HapMap Consortium (2007)
- the priors of K and \underline{T} are similar to mBPCR (Rancoita et al. (2009)):

$$P(\underline{T} = \underline{t} | K = k) = \binom{n-1}{k-1}^{-1}, \quad \underline{t} \in \mathbb{T}_{k,n}$$
$$P(K = k) = \frac{k_{\max} + 1}{k_{\max}} \frac{1}{k(k+1)}, \quad k \in \mathbb{K} = \{1, \dots, k_{\max}\}$$

- $P(Z_p = z)$ derived from the mBPCR estimated profile of CN data (we need to map the continuous \log_2 ratio values into the classes of CN aberrations):

$$\begin{aligned} P(Z_p = 2) &= P(M_p \geq \hat{\mu}_4 + 3\hat{\sigma}_4 | cn) \\ P(Z_p = 1) &= P(\hat{\mu}_2 + 3\hat{\sigma}_2 < M_p \leq \hat{\mu}_4 + 3\hat{\sigma}_4 | cn) \\ P(Z_p = 0) &= P(\hat{\mu}_2 - 3\hat{\sigma}_2 < M_p \leq \hat{\mu}_2 + 3\hat{\sigma}_2 | cn) \\ P(Z_p = -1) &= P(\hat{\mu}_1 - 3\hat{\sigma}_1 < M_p \leq \hat{\mu}_2 - 3\hat{\sigma}_2 | cn) \\ P(Z_p = -2) &= P(M_p \leq \hat{\mu}_1 - 3\hat{\sigma}_1 | cn), \end{aligned}$$

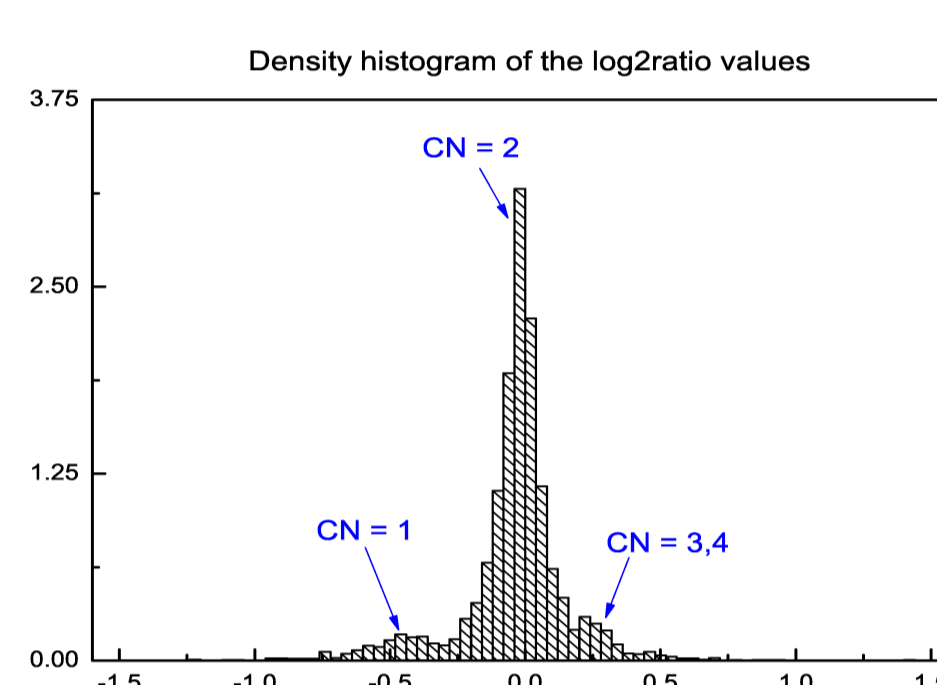
where:

cn = all the information regarding the copy number data

M_p = CN value in the p^{th} interval, $M_p \sim \mathcal{N}(\hat{m}_p, \hat{V}_p)$

(\hat{m}_p, \hat{V}_p) = posterior mean and variance of M_p estimated by mBPCR

$(\hat{\mu}_{cn}, \hat{\sigma}_{cn}^2)$ = estimated mean and variance of the normal distribution corresponding to $CN = cn$



From the model, the posterior of \tilde{W} is:

$$p(\tilde{w} | \underline{y}, \underline{t}^0, k_0) \propto \prod_{p=1}^{k_0} \prod_{i=t_{p-1}^0+1}^{t_p^0} \sum_{x \in \mathbb{X}} p(y_i | X_i^N = x, w_p) P(X_i^N = x | w_p),$$

The estimation

- Method 1 (similar to mBPCR):

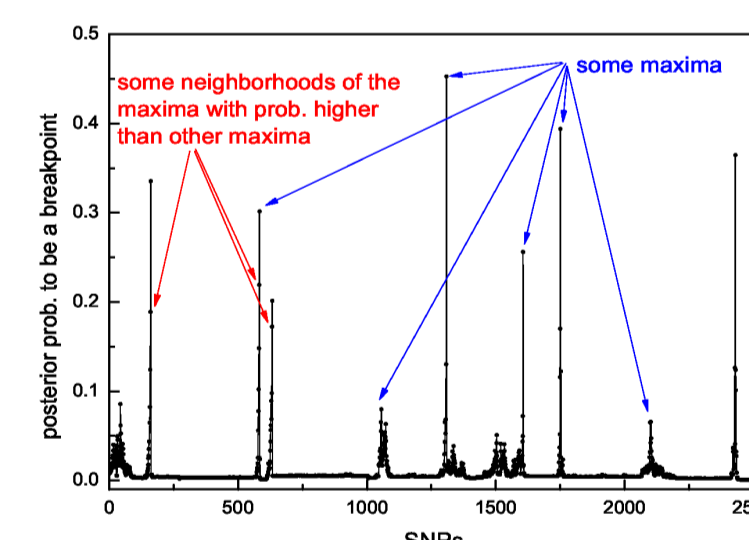
$$\hat{K}_{01} = \arg \max_{k \in \mathbb{K}} p(k | \underline{Y}, cn),$$

$$\hat{T}_{BinErrAk} = \arg \max_{\underline{t} \in \mathbb{T}_{k,n}} \sum_{q=1}^{k-1} \sum_{p=1}^{k_0-1} \delta_{t_p^0, t_q^0} | \underline{Y}, cn$$

$$\hat{W}_p = \arg \max_w P(W_p = w | \underline{Y}, \hat{t}, \hat{k}, cn), \quad p = 1, \dots, k$$

- $\hat{T}_{BinErrAk}$ consists of the \hat{k}_{01} positions which have the highest posterior probability to be a breakpoint p_i

⇒ problem: we could take some points in the neighborhood of the higher maxima of p and not some maxima with a lower probability



- Method 2: estimate the number of the segments and the breakpoints with, respectively, the number of peaks and the locations of their maxima (\underline{W} estimated as previously)

- It uses two thresholds: one for the determination of the peaks (thr_1) and one for the definition of the values close to zero (thr_2).

⇒ corresponding estimators $\hat{K}_{Peaks,thr_1,thr_2}$ and $\hat{T}_{Peaks,thr_1,thr_2}$ (the method is denoted with (thr_1, thr_2))

- Paired thresholds selected on the basis of results obtained on simulations: (01, 01), (mad, 01), (01, mad), where

$$01 = \max(0.01, \text{quantile of } \underline{p} \text{ at } 0.95)$$
$$mad = \text{median}(\underline{p}) + 3 * \text{mad}(\underline{p})$$

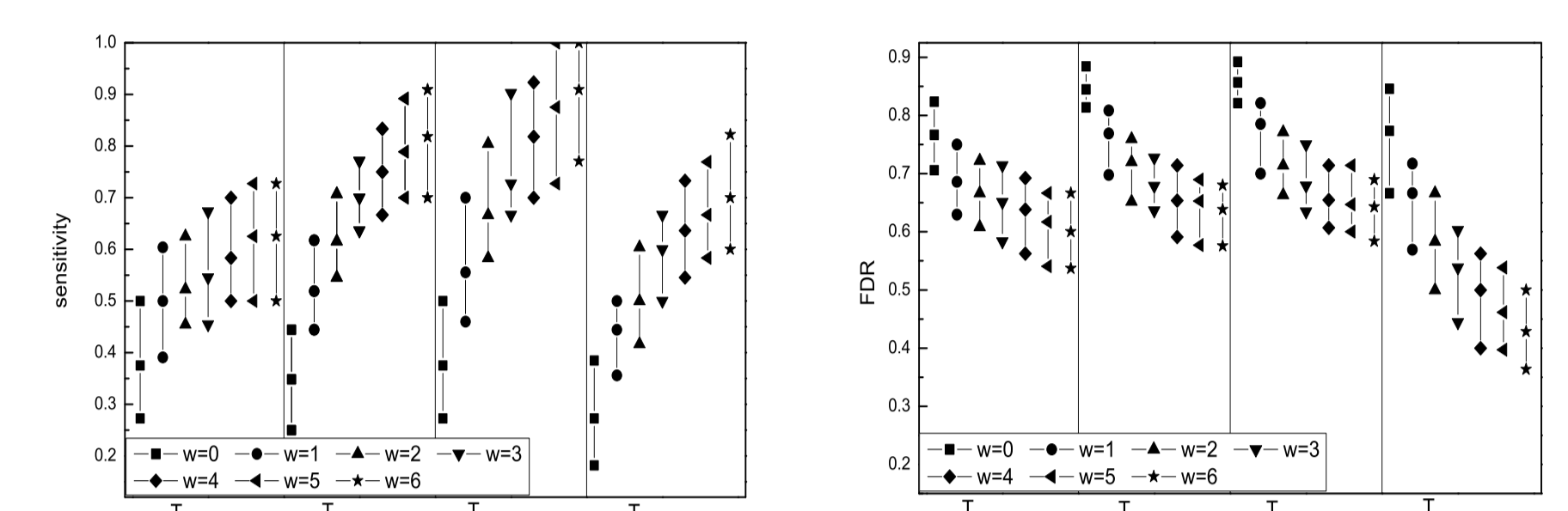
Some results on simulations

- Aberrations not considered in the simulations:
 - gain (because it does not influence the genotype detection)
 - IBD/UPD (difficult to simulate realistically)

- Simulated dataset (100 samples with fixed k_0 and \underline{t}^0): each sample is a raw profile coming from the prior definition of \underline{X}^N given by the annotation file for the SNPs of chr. 22 in the Affymetrix GeneChip Mapping 250K Array ($n = 2520$) and the following prior definition of \underline{Z} ($P(Z_p = z) =: q^z$)

	segment														
	I	II	III	IV	V	VI	VII	VIII	IX	X	XI	XII	XIII	XIV	XV
q^1	0	0.1	0	0.1	0.5	0.1	0	0	0.1	0.5	0	0.1	0.5	0.1	0
q^0	0.1	0.6	0.1	0.6	0.4	0.6	0.1	0.1	0.6	0.4	0.1	0.6	0.4	0.6	0.1
q^{-1}	0.6	0.3	0.6	0.3	0.1	0.3	0.6	0.4	0.3	0.1	0.6	0.3	0.1	0.3	0.6
q^{-2}	0.3	0	0.3	0	0	0	0.3	0.5	0	0	0.3	0	0	0	0.3

- Some results on breakpoint estimation:



⇒ Method 2 has higher sensitivity and similar or lower FDR.

- Some results CN aberration estimation (- best result, - worst result):

method	sum 0-1 err	SSQ	$\sqrt{SSQ/n}$
method 1	421.79	1226.59	0.70
(01, 01)	109.39	286.15	0.34
(01, mad)	109.39	286.15	0.34
(mad, 01)	111.75	283.77	0.34

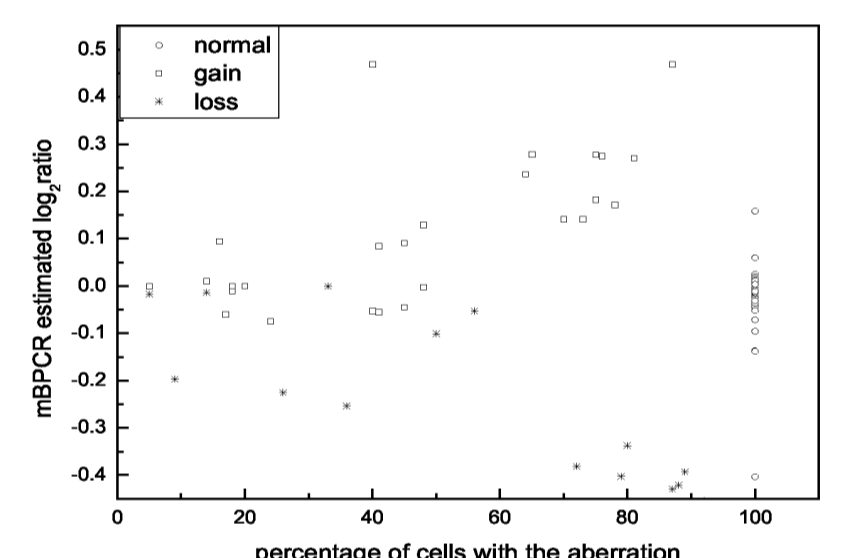
method	sensitivity				FDR			
	Z=2	Z=0	Z=-1	Z=-2	Z=2	Z=0	Z=-1	Z=-2
method 1	0.681	0.932	0.968	0.555	0.017	0.047	0.306	0.025
(01, 01)	0.896	0.983	0.961	0.946	0.043	0.031	0.068	0.020
(01, mad)	0.896	0.983	0.961	0.946	0.043	0.031	0.068	0.020
(mad, 01)	0.889	0.984	0.963	0.942	0.038	0.026	0.075	0.023

⇒ Method 2 best estimates the profile (best paired threshold: (01, 01), (01, mad)).

Applications on real data

- Data: paired samples of patients affected by chronic lymphocytic leukemia (CLL), which then transformed in diffuse large B-cell lymphoma (DLBCL) (Bertoni et al. (2008)). Of two patients, we had three samples.

- detectable CN aberrations = the ones born by at least 60% of cells in the sample



- Evaluation of the estimation of the CN aberrations: comparison with the estimated CN of some genomic regions with FISH (fluorescent in situ hybridization), which gives also the percentage of cells bearing the aberration

- 15/17 detectable aberrations found by all estimators
- 3/26 not detectable aberrations found by all estimators and another by (01, 01) and (01, mad) with $p_{upd} = 10^{-3}$ and (mad, 01) with $p_{upd} = 10^{-4}$
- in only 2/90 normal segments, all estimators discovered an aberration
- Remark: a slightly discordance between the 2 techniques is possible, because the samples used are not exactly the same

- Evaluation of the IBD/UPD region detection: comparison of the regions found in the 3 samples of 2 patients

Patient 1:	types of regions	$p_{upd} = 10^{-4}$			$p_{upd} = 10^{-3}$			
		01, 01	01, mad	mad, 01	01, 01	01, mad	mad, 01	
distinct (total)	equal (%)	0.79	0.79	0.78	0.78	0.78	0.77	
	equal in 2 samples (%)	0.15	0.15	0.20	0.15	0.15	0.18	
	overlapping (%)	0.03	0.03	0.01	0.02	0.02	0.03	
	validated (%)	0.98	0.98	0.98	0.95	0.95	0.98	
	remaining (%)	0.02	0.02	0.02	0.05	0.05	0.02	
	% of remaining < 1Mb	0.80	0.80	0.88	0.93	0.92	1.00	
	Patient 2:	distinct (total)	441	441	454	580	580	618
		equal (%)	0.21	0.21	0.25	0.19	0.19	0.24
		equal in 2 samples (%)	0.02	0.02	0.03	0.03	0.03	0.02
overlapping (%)		0.50	0.50	0.47	0.51	0.51	0.50	
validated (%)		0.73	0.73	0.74	0.74	0.74	0.76	
remaining (%)		0.27	0.27	0.26	0.26	0.26	0.24	
% of remaining < 1Mb		0.88	0.88	0.89	0.91	0.91	0.93	

⇒ the 3 estimators behaved similarly and equally well on real data

Summary and conclusions

- Our method is a new algorithm for the joint estimation of CN events and IBD/UPD regions, which takes into account the errors in the genotyping measurements of microarrays, due to the aberrations affecting the CN.
- Differently from the only other method present in literature (i.e., Scharp et al. (2008)), it considers all the CN events biologically relevant.
- The goodness of our model is supported by the results obtained on simulated and real data.